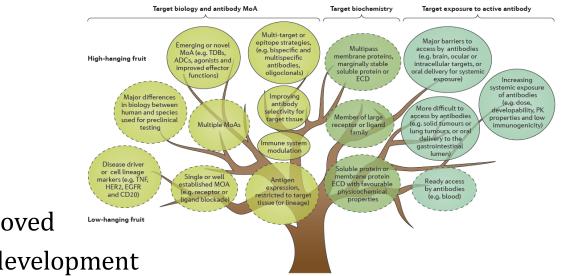


Human Antibody Identification Services: Exiris Library



Monoclonal Antibodies: the next challenges



- Mature field
- > 70 mAbs approved
- > 600 mAbs in development

Nat Rev Drug Discov. 2018 Mar;17(3):197-223.

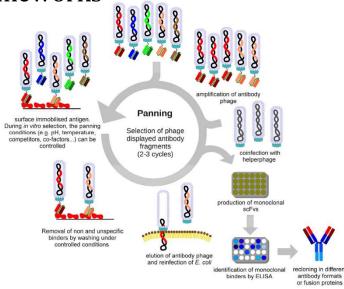
- Challenges:
 - Novel MOAs: agonist mAbs, mAbs inhibiting enzyme functions
 - Multispecific mAbs



A high complexity, synthetic, phage-displayed antibody library

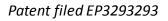
Features:

- High complexity: 1.34×10^{10}
- scFV format, fully human
- Highly validated VH3/VK1 frameworks



Advantages:

- Maximize probability of identifying high affinity binders
- Maximize identification of mAbs with good developability
- Facile identification of low abundance clones with novel biologic properties
- Straightforward development of bispecific binders



Complexity of 1.34×10^{10}

Complexity	Probability to identify a binder with Kd <1 nM
10 ⁷	1%
10 ⁸	9.6 %
10 ⁹	63.2 %
10 ¹⁰	99.9%

The probability **P** of identifying a ligand within a library that has an affinity lower than the threshold value **p** depends on the number of different ligands **N**, present in the library according to the formula $P = 1 - e^{-Np}$

Complexity> 10⁹ is needed to have high probability of identifying high affinity binders

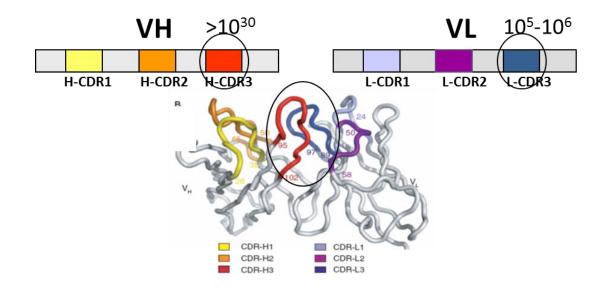


Library design principles

- Library diversity only in HC-CDR3
 - Major determinant of antibody diversity
- Focus on HC-CDR3 loop lengths frequently present in therapeutic mAbs
 - Increase probability of success during development
- Reduce combinatorial bias (oversampling of short HC-CDR3, undersampling of long HC-CDR3)
 - Adjust percent fraction of loop lengths and reduce aa diversity going from shorter to longer HCDR3



Diversity only in HC-CDR3

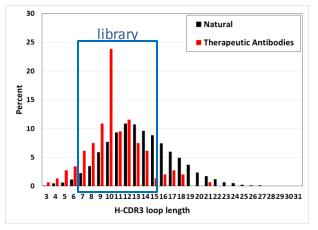


HC-CDR3 is the major contributor to antibody diversity



Specific loop length distribution

HC-CDR3 loop length distribution Natural (black), Therapeutic mAbs (red)

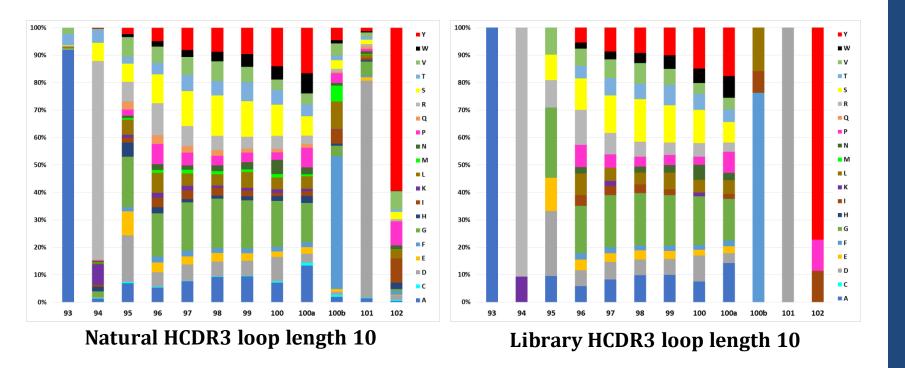


Germline HC-CDR3 length distribution is broad. Therapeutic antibodies show a pronounced maximum at HC-CDR3 loop length 7-14 possibly reflecting good physicochemical properties (solubility, no aggregation, etc) and good expression properties

Exiris library loop length distribution based on natural distribution but especially focused on the main portion of therapeutic mAbs loop lengths (7-15 aa)

H.)

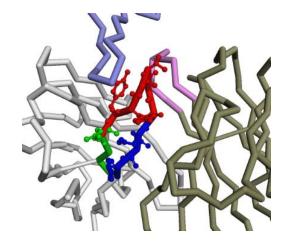
Non-random amino acid distribution



In each HC-CDR3 loop position we incorporated residues preferentially found in nature, but rare variants were excluded

Ex

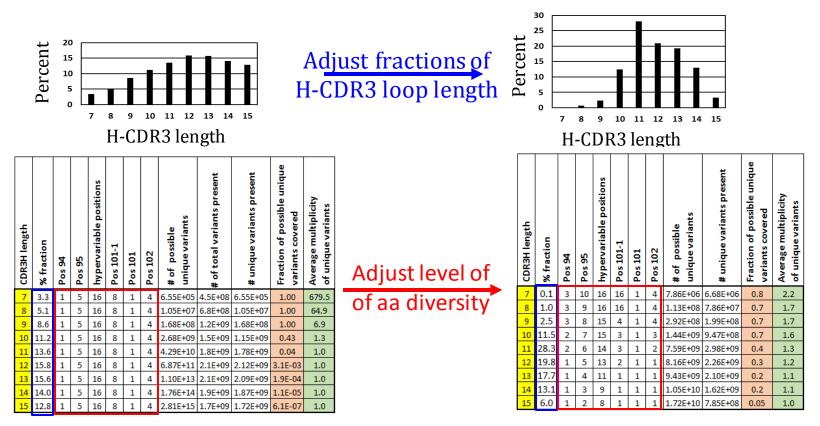
Non-random amino acid distribution



The initial (**dark blue**) and final (**green**) segments of the HC-CDR3 loop do not in general directly contribute to antigen recognition. Direct contacts with the antigen (**light blue**) are mostly mediated by the central segment of the HC-CDR3 (**red**).

In each HC-CDR3 loop position the initial and final parts were kept relatively constant; in longer loops less variability was introduced in order to cover a larger fraction of possible theoretical variants

Non-random amino acid distribution



Both percentage of each loop length and aa composition in each loop position were adjusted in order to minimize redundancy of shorter loops and maximize coverage of longer loops

Ex

Final design

- H-CDR3, complexity 1.34 x 10¹⁰
- Exclude Cys, His, Met, Gln
- Focus on loop distribution found in therapeutic mAbs (Loop lengths: 7-15)
- Naturally found amino acid frequency, exclude rare variants
- Modulate aa diversity for each loop to maximize coverage

			number of																						
			different																					Actual	Redundancy
			amino																				Fraction of	number	(number of
	fraction of	number	acids at									_			-						Number of	Actual	theoretically	of variants	times each
	total loops	of hyper-	hyper-	~	8	35	96	97	8	66	10	100a	100b	100c	100d	100e	100f	100g	101	102	theoretically	number	possible variants	present	variant
H-CDR3	in library	variable	variable	s 93	Pos 9	Pos 9	Pos 9	os 9	Pos 9	Pos 9	Pos 1	os 1	os 1	Pos 1	Pos 1	possible	of clones	actually present	(Poisson	is present					
length	(%)	positions	positions	Po	ă	ă	ă	ă	ă	ă	ă	ă	ă	ď	ď	ă	ă	ă	ă	ă	variants	present	(Poisson estimate)	estimate)	on average)
7	0.1	3	16	1	3	10	16	16	16	16									1	4	7.86E+06	1.49E+07	0.85	6.68E+06	2.2
8	1.0	4	16	1	3	9	16	16	16	16	16								1	4	1.13E+08	1.34E+08	0.69	7.86E+07	1.7
9	2.5	5	15	1	3	8	15	15	15	15	15	4							1	4	2.92E+08	3.35E+08	0.68	1.99E+08	1.7
10	11.5	6	15	1	2	7	15	15	15	15	15	15	3						1	3	1.44E+09	1.55E+09	0.66	9.47E+08	1.6
11	28.3	7	14	1	2	6	14	14	14	14	14	14	14	3					1	2	7.59E+09	3.79E+09	0.39	2.98E+09	1.3
12	19.8	8	13	1	1	5	13	13	13	13	13	13	13	13	2				1	1	8.16E+09	2.65E+09	0.28	2.26E+09	1.2
13	17.7	9	11	1	1	4	11	11	11	11	11	11	11	11	11	1			1	1	9.43E+09	2.37E+09	0.22	2.10E+09	1.1
14	13.1	10	9	1	1	3	9	9	9	9	9	9	9	9	9	9	1		1	1	1.05E+10	1.76E+09	0.15	1.62E+09	1.1
15	6.0	11	8	1	1	2	8	8	8	8	8	8	8	8	8	8	8	1	1	1	1.72E+10	8.04E+08	0.05	7.85E+08	1.0

Outcome: no or very low redundancy, excellent representation of intermediate and long loops **Greatly improved diversity**



Library validation: (1) Quality control

NGS of 2773807 sequences:

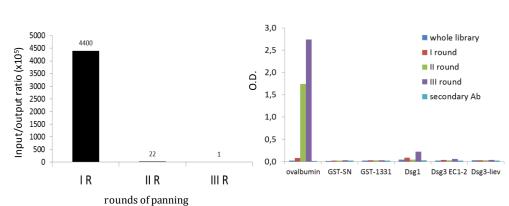
- Correct amino acid composition
- ✓ Relative frequency of amino acids as designed
- ✓ 94% of clones with inserts
- \checkmark 6-7% of clones with insertions or deletions
- ✓ 88% correct sequences
- \checkmark very few duplications, in line with design

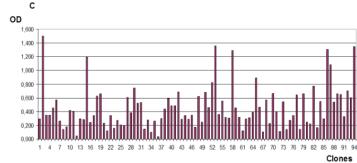


Library validation: (2) Panning against test antigen (ovalbumin)

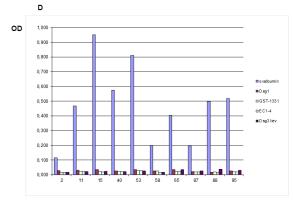
1. panning

2. Individual clones





3. Selected clones



4. Sequencing

CLONE	V _H (DP47)
2	CAKVVTGVLWGFDYW
11	CAKD
58	CAKDWGYFDYW
15	CAKDFGRGYGYFDYW
65	CAKDFRSGYGYFDYW
40	CAQDVRRGCGYFDYW
53	CAKDVARGYGYFDYW
95	CAKDVWRGYGYFDYW
88	CAKDVGRGVGYFDYW
87	CAKVVGGVLYAFDYW

Library validation: (2) Panning against panel of test antigens – summary results

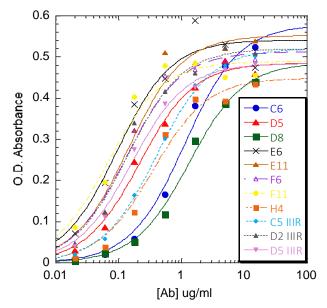
	Number of rounds	# analyzed clones	# positive clones	% anti-target positive clones
BSA	3	47	89	53%
OVA	3	71	94	76%
GST	3	20	90	22%
DSG1	3	81	94	86%
FGFR4	3	73	77	95%
target 1	3	49	49	100%
target 2	3	135	135	100%

	Specificity (vs other targets)	Sequenced clones	# of unique sequences
BSA	12/12	5	1
OVA	10/10	10	10
GST	15/15	10	8
DSG1	10/10	10	10
FGFR4	16/20	10	4
target 1	16/16	10	8
target 2	75/135	75	31



Project 1: selection of high affinity binders (IgG1) against recombinant cell surface receptor

Kd (nM) of identified IgGs (ELISA)



Antibody	Kd (nM)
C6	7,5
D5	1,4
D8	8,8
E6	0,6
E11	0,9
F6	0,9
F11	0,4
H4	2,3
C5 IIIR	2,6
D2 IIIR	0,9
D5 IIIR	1,1

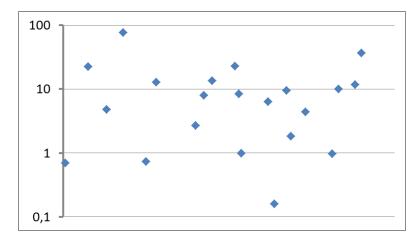
80% productive and functional IgG 100% with Kd < 10 nM

Best affinity: Worst affinity: 400 pM 8.8 nM



Project 2: selection of high affinity binders (IgG1) against "difficult" target

Kd (nM) of selected IgGs (ELISA)

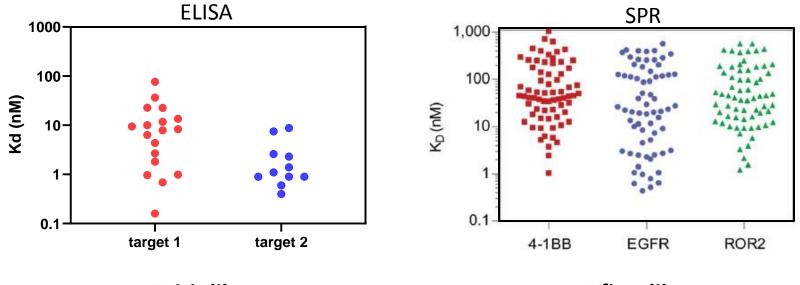


Kd < 1 nM:	24%
Kd < 5 nM:	43%
Kd < 10 nM:	67%
Kd< 25 nM:	90%

Best affinity: Worst affinity: 160 pM 76 nM



Comparison between Exiris library and competition



Exiris library

Pfizer library

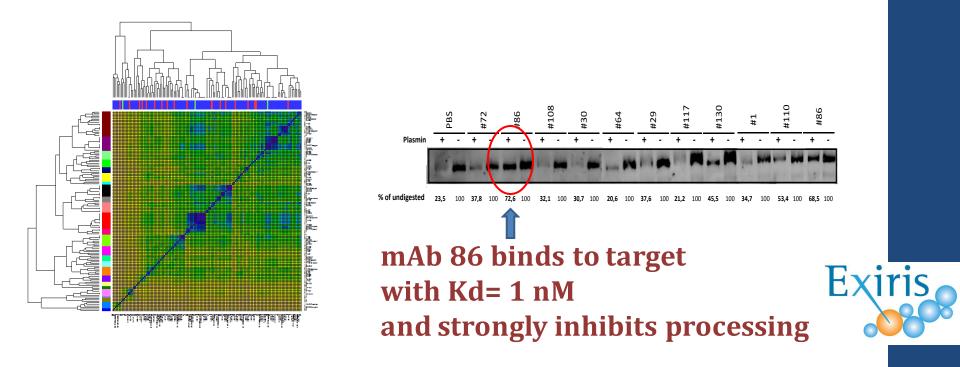
(Van Blarcom et al. MABS, 2018)

Superior performance of Exiris library

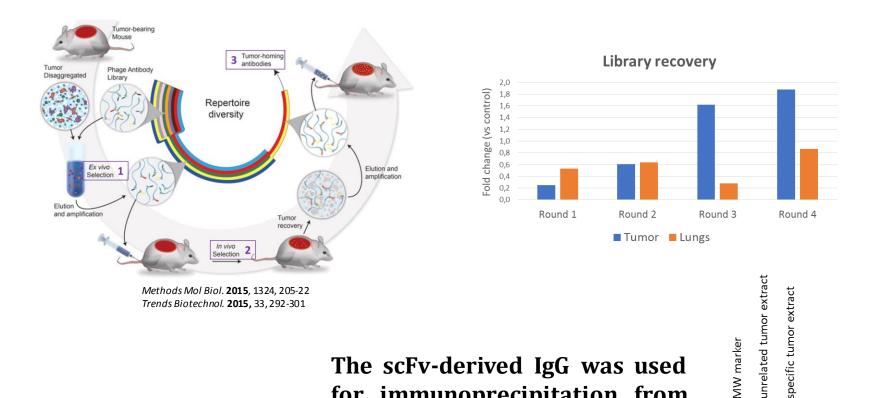


Project 3: selection of antibodies with specific biological function (inhibitors of proteolytic processing)

Extracellular target. Proteolysis releases cytokine involved in pathologic processes. So far only polyclonal Ab mixtures were reported to inhibit processing. Exiris library screen of 200 binders revealed 19 sequence families. Representative members were cloned, expressed and purified.



Project 4: In vivo selection



The scFv-derived IgG was used for immunoprecipitation from tumor extracts and led to the identification of the target protein by mass spectrometry

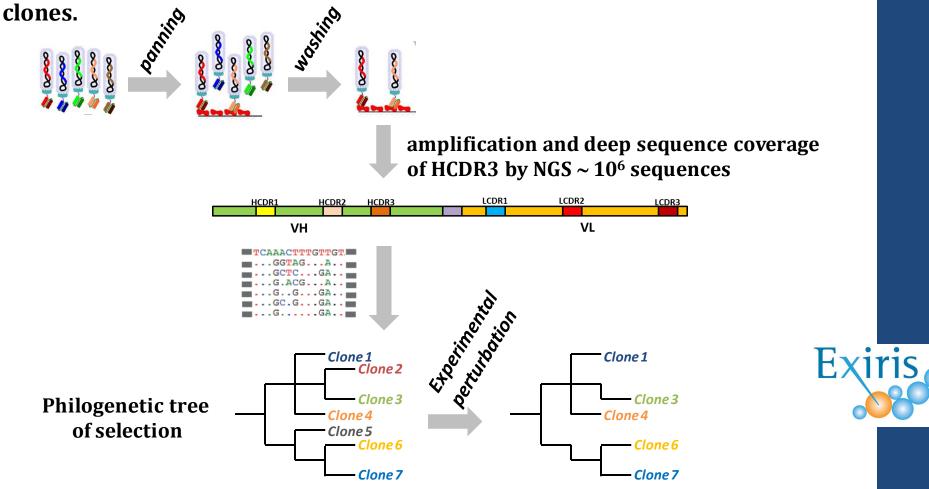
Identification of specific tumor-homing phages



MW marker

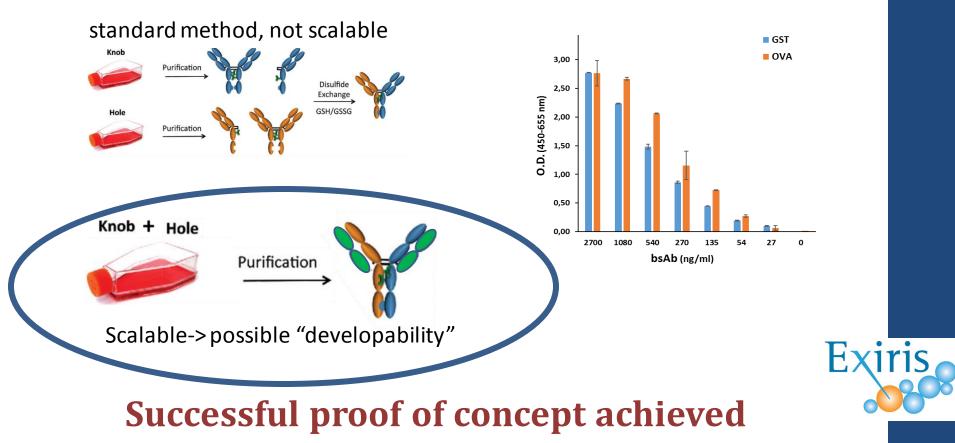
Advantages of the Exiris library: (1) straightforward ID of low abundance clones

During standard library selections high affinity binders are often missed, even by sampling > 5000 clones. Following selection by NGS of a high complexity "H-CDR3-only" –library maximizes the likelihood of identifying low abundance



Advantages of the Exiris library: (2) straightforward access to bispecifics

An "H-CDR3-only" –library allows to rapidly generate bispecific mAbs using the "knobs into holes" technique and a single host cell production.



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